

M. C. E. Society's Abeda Inamdar Senior College

Of Arts, Science and Commerce, Camp, Pune-1 (Autonomous) Affiliated to Savitribai Phule Pune University NAAC accredited 'A' Grade

SR. NO.	COURSE CODE	COURSE NAME
1	23SBMB51MM	Immunology - I
2	23SBMB52MM	Enzymology
3	23SBMB53MM	Bacterial Genetics
4	23SBMB54MM	Practical course: Enzymology and Genetics
5	23SBMB55MM	Practical course: Industrial and Agricultura Microbiology
6	23SBMB51MEA	Fermentation Technology - I
7	23SBMB51MEB	Marine Microbiology
8	23SBMB52MEA	Agricultural Microbiology
9	23SBMB52MEB	Dairy Microbiology
10	23SBMB51VS	Practical course: Immunology



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Of Arts, Science and Commerce, Camp, Pune- 1 (Autonomous) Affiliated to Savitribai Phule Pune University NAAC accredited 'A' Grade

T.Y.B.Sc. Microbiology

(Autonomy NEP 2023 Pattern)

Course/ Paper Title	Immunology I
Course Code	23SBMB51MM
Semester	V
No. of Credits	2

Aims & Objectives of the Course

Sr. No.	Objectives
1.	To understand a comprehensive understanding of the types of immunity
2.	To understand the importance of structure and function of primary and secondary lymphoid organs
3.	To understand the components and mechanisms involved in innate immunity and non-specific defense, which are crucial for the body's initial response to pathogens
4.	To gain practical skills and knowledge about various antigen-antibody interaction techniques in clinical settings and enhancing their ability to diagnose and monitor diseases effectively.
5.	To understands the concepts of immunohematology and their applications in clinical diagnostics

Sr. No.	Learning Outcome
1.	Students will be acquainted with the concepts of immunity and organs involved
	in it.
2.	Students will become aware about the role of immune cells in different diseases
3.	Students will understand the clinical tests and their applications in disease diagnosis.

Unit No.	Title with Contents	No. of Lectures
Credit I	Immune System: Structure and Function	15
1.	a. Immunity – Definition & Classification of Immunity- Innate &	2
	Acquired Immunity, Active and passive immunity, Humoral & cell	
	mediated Immunity	1
	b. Three lines of defence mechanisms	2
	c. Cells of Immune system: Hematopoiesis, Structure, classification,	
	properties and function of Stem cell, T cell, B cell, NK cell,	
	Macrophage, Neutrophil, Eosinophil, Basophil, Mast cell,	2
	Dendritic cell.	2
	d. Lymphoid Organs (primary and secondary)- location,	
	classification & functions of -Bone Marrow, Thymus, Lymph	
	Node, Spleen, GALT, MALT, CALT	
2.	Antigen - Definition, concept of immunogen, epitopes, types of antigens- soluble and particulate antigens, autoantigens, Isoantigens, factors affecting immunogenicity. Concepts of haptens, carriers and adjuvants	1
3.	Antibodies/Immunoglobulin - Definition, structure of typical immunoglobulin, classes and biological functions of immunoglobulin, light and heavy chain domains	2
4.	Principles of antigen and antibody interactions- lattice hypothesis and	1
	zone phenomenon Antigen-antibody Reactions – i. Precipitation	4
	ii. Agglutination	
	iii. IF	
	iv. ELISA	
	v. RIA	
	vi. FACS	
Credit II	Antigen- Antibody Interactions, Major Histocompatibility	15
	Complex, Transplantation and Immunity and	
	Immunohematology	

1.	Immune response: Cellular and Humoral	
	a. Cellular components: Phagocytic cells – PMNL, macrophages (reticulo- endothelial cell system) and dendritic cells	1
	b. Humoral components: complement, cytokinins, inflammatory mediators	1
	c. Non-specific mechanisms of defense	
	d. Phagocytosis - oxygen dependent and independent systems	2
	e. Complement activation - Classical, Alternative and lectin	
	pathway	
	f. Inflammation - cardinal signs, mediators, vascular and cellular	
	changes	
2.	Major Histocompatibility Complex:	1
	Structure of MHC in man and mouse	$\frac{1}{2}$
	Structure and functions of MHC class–I and class–II molecules	
	MHC antigen typing (microcytotoxicity, mixed lymphocyte	
	reaction and molecular typing)	
3.	Transplantation and Immunity:	1
	Types of Grafts, Allograft rejection mechanisms Prevention of	1
	allograft rejection Shelf-life determination	
4.	Immunohematology	3
	 a. ABO and Rh blood group systems b. Bombay blood group c. Biochemistry of blood group substances d. Inheritance of ABH antigens e. Medico- legal applications of blood groups 	
5.	Immunomodulatory Approaches in Fungal and Cancer Diseases a. Concept b. Significance in treating infection and cancer	2

References: Immunology- I

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T.Y.B.Sc. Microbiology

(Autonomy NEP 2023 Pattern)

Course/ Paper Title	Enzymology
Course Code	23SBMB52MM
Semester	V
No. of Credits	2

Aims & Objectives of the Course

Sr. No.	Objectives
1.	To understand methods of active site determination, role of enzymes and its cofactors in microbial physiology.
2.	To learn to perform enzyme assay, purification and quantification of enzymes activity, enzyme kinetics in terms of initial, final velocity, mathematical expression of enzyme kinetic parameters.
3.	To correlate regulation of metabolism at enzymatic levels and apply methodology for commercial applications of enzymes.

Sr. No.	Learning Outcome
1.	Students will be acquainted with the methods of active site determination, role of enzymes and its cofactors in microbial physiology.
2.	Students will become aware about the concept enzyme assay, purification and quantification of enzymes activity, enzyme kinetics in terms of initial, final velocity and mathematical expression of enzyme kinetic parameters.
3.	Students will understand the significance of regulation of metabolism at enzymatic levels and apply methodology for commercial applications of enzymes.

Unit No	Title with Contents	No. of Lectures
Credit I	Enzymes:	15
1	Structure of enzymes: a. Methods to determine amino acid residues at active site (Physical method e.g. x-ray crystallography and chemical methods such as trapping of ES complex and use of pseudo- substrate)	03
	b. Role of vitamins in metabolism: Occurrence, Structure and Biochemical functions of the following: i) Nicotinic Acid (Niacin) and the Pyrimidine nucleotides. ii) Riboflavin (Vitamin B2) and the Flavin nucleotides	02
2	Enzyme assays: a. Principles of enzyme assays and calculation of enzyme unit, specific activity	01
	b. Enzymes assays with examples by: i. Spectrophotometric methods ii. Radioisotope assay	02
3	3. Principles and Methods of Enzyme purification: a. Methods of cell fractionation b. Principles and methods of enzyme purification:	01
	i. Based on molecular size	02
	ii. Based on charge	01
	iii. Based on solubility differences	01
	iv. Based on specific binding property and selective adsorption	01
	c. Construction of enzyme purification chart	01
Credit II	Enzyme Kinetics, metabolic regulation and Immobilized Enzymes:	15
1	Enzyme Kinetics:	
	a. Concept of initial velocity	01
	b. Michaelis Menton equation for the initial velocity of single	
	substrate enzyme catalyzed reaction. Brigg's Haldane modification of Michaelis Menton equation. Michaelis Menton plot, Lineweaver and Burk plot. Definition with significance of Km, Ks, Vmax.	05
2	Metabolic Regulations:	
	a. Enzyme compartmentalization at cellular level	01
	b. Allosteric enzymes	01
	c. Feedback mechanisms	01
	d. Covalently modified regulatory enzymes (Glycogen phosphorylase) e. Proteolytic activation of zymogens	01 01
	f. Isozymes - concept and examples	01
	g. Multienzyme complex e.g. Pyruvate dehydrogenase complex (PDH)	01
3	Immobilization of enzymes:	02
	Concept, methods of immobilization and applications	

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T.Y.B.Sc. Microbiology

(Autonomy NEP 2023 Pattern)

Course/ Paper Title	Bacterial Genetics
Course Code	23SBMB53MM
Semester	\mathbf{V}
No. of Credits	2

Aims & Objectives of the Course

Sr. No.	Objectives
1.	To understand the central dogma of molecular biology.
2.	To know basic biological life processes like replication, transcription and translation.
3.	To learn basic gene transfer and mapping techniques.
4.	To know basic recombination mechanisms at the molecular level.

Sr. No.	Learning Outcome
1.	To exhibit a knowledge base in Bacterial Genetics.
2.	To understand the central dogma of Molecular Biology.
3.	To construct genetic map of bacteria.
4.	To get introduced to the concept of recombination.

Unit No	Title with Contents	No. of
		Lectures
Credit I	DNA REPLICATION, CENTRAL DOGMA AND GENE	15
	EXPRESSION	
	1. Process of prokaryotic DNA replication	05
	a. Concept of Single replicon.	03
	b. Bidirectional movement of replication fork, Ori C gene	
	c. Pre-priming and Priming reactions.	
	d. DNA polymerases, DNA synthesis of leading & lagging strand	
	synthesis. Okazaki fragments.	
	e. Termination-Ter sequences, Tus protein	
	2. Transcription	05
	a. Concept of central dogma	03
	b.Structure of promoter sequences.	
	c. Structure and function of RNA polymerase.	
	d. Steps of transcription: Initiation, Elongation and termination.	
	e.Gene Regulation and Operon Concept (Eg-Lac Operon)	
	3. Translation	05
	a. Structure and roles of m-RNA, t-RNA, rRNA and ribosome in	
	translation.	
	b. Role of Aminoacyl t-RNA synthetase in tRNA charging.	
	c. Steps in translation: Initiation, elongation, translocation and	
	termination of protein synthesis.	
Credit II	Gene transfer in bacteria and mapping	15
	techniques.	
	1. Gene transfer by Transformation	4
	a. Discovery of Transformation	
	b. Natural transformation Systems-Streptococcus pneumonia and	
	Haemophilus influenzae.	
	c. Factors affecting transformation process (Competence development,	
	Size of DNA, Concentration of DNA)	
	d. Mapping of genes by co-transformation	

2. Gene transfer by Conjugation	4
a. Discovery of Conjugation	
b. Properties of F plasmid, F ⁺ ,F ⁻ , Hfr and F' strains	
c. Process of conjugation between F ⁺ and F ⁻ , Hfr and F ⁻	
d. Mapping by interrupted mating experiment	
3. Gene transfer by Transduction	4
a. Discovery of Transduction	
b. Generalized transduction mediated by phage P22	
c. Specialized transduction mediated by lambda phage.	
d. Mapping of genes by co-transduction	
4. An introduction to Recombination	3
a. Concept of genetic recombination and its types.	
b. Concept of recombination mapping: Map unit, recombination	
frequency, recombination frequency percentage.	
c. Concept of Holliday model of recombination, Role of Rec and Ruv	
proteins in homologous recombination.	

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- 4. University of North Carolina's Biosafety Guidelines (Principles, Risk assessment, Biosafety levels, Guidelines):

https://ehs.unca.edu/laboratory-safety/biological-safety/ http://www.informatics.jax.org/silver/chapters/7-1.shtml



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T.Y.B.Sc. Microbiology 2025-26 (NEP-2023)

Course/ Paper Title	Practical course: Enzymology and Genetics
Course Code	23SBMB54MM
Semester	V
No. of Credits	2

Aims & Objectives of the Course

Sr.	Objectives
No.	
1.	To make students aware about the principle and working of
	colorimeter and spectrophotometer along with estimation of
	biological molecules.
2.	To make students aware about basics of Enzymology along with
	production, assay and precipitation of Amylase.
3.	To make them understand basic chromatography techniques with an
	example of Paper Chromatography.
4.	To introduce basic techniques in Molecular Biology like plasmid
	DNA isolation, estimation and purity determination

Sr. No.	Learning Outcome
1.	Students will learn the principle and working of colorimeter and
	spectrophotometer along with estimation of biological molecules.
2.	Students will learn basics of Enzymology along with production, assay
	and precipitation of Amylase.
3.	Students will be acquainted with concept, principle and basic
	chromatography techniques with an example of Paper Chromatography
4.	Students will understand principle and methodology of plasmid DNA
	isolation, estimation and purity determination

Expt. No.	Topics	No. of Practicals
1	Determination of absorption spectra and molar extinction co-efficient of two	2
	different dyes (by colorimetry /spectrophotometry)	
2	Estimation of reducing sugar from natural sources by DNSA method	1
3	Estimation of proteins from natural sources by Folin Lowry method	1
4	Lab scale production of amylase using isolates	1
5	Assay of Amylase Enzyme	1
6	Precipitation of amylase from fermentation broth by salt &	2
	Determination of specific activity of crude and purified amylase	
7	Separation and Identification of amino acids from the mixture by paper chromatography.	1
8	Isolation of Plasmid DNA from Bacteria	1
9	Estimation of DNA by Diphenylamine method.	1
10	Determination of purity of DNA preparation and its quantification.	1
	Estimation of DNA by UV spectrophotometric methods at 260 nms. Purity	
	checks of DNA by 260 / 280 ratio.	
11	Bacterial Artificial transformation & Competence development in <i>E coli</i> using	2
	Calcium Chloride method.	
12	Bacterial Conjugation(Demonstration)	1
	TOTAL	15



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T.Y.B.Sc. Microbiology (Autonomy NEP 2023 Pattern)

Course/ Paper Title	Practical course: Industrial and Agricultural
	Microbiology
Course Code	23SBMB55MM
Semester	V
No. of Credits	2

Aims & Objectives of the Course

Sr.No.	Objectives
1.	To make students aware about operation of laboratory scale fermenter
2.	To make them understand importance of sterility checking of injectables
3.	To introduce concept of bioassay by different methods
4.	To make them understand importance of microorganisms in plant pathology and agriculture

Sr. No.	Learning Outcome
1.	Students will learn about the use of laboratory scale fermenter
2.	Students will learn to check sterility of injectables and get acquainted with different types of methods used for bioassay
3.	Students will learn isolation and identification of plant pathogens
4.	Students will understand importance of microorganisms with respect to soil fertility and plant growth

Expt.	Topics	No. of
No.		Practicals
1	Media preparation and sterilization of laboratory scale fermenter	2
2	Sterility Testing of pharmaceuticals (non-biocidal injectables)	1
3	Minimum inhibitory concentration and minimum bactericidal concentration of antibacterial compounds (MIC and MBC)	2
4	Antibiotic and growth factor assay (agar gel diffusion technique)	2
5	Isolation and identification of <i>Xanthomonas</i> spp. from citrus canker	1
6	Isolation of Aspergillus niger from black rot of onion	1
7	Collection of plant disease specimens and study of symptoms/ Project based on digital record of plant diseases (Group Activity)	2
8	Isolation of PGPR with phosphate solubilization potential	1
9	Validation of commercial formulations of bioinoculants based on BIS standards,	1
10	Pot studies to check effect of bioinoculants on plant growth	2
	TOTAL	15

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T.Y.B.Sc. Microbiology

(Autonomy NEP 2023 Pattern)

Course/ Paper Title	Fermentation technology I
Course Code	23SBMB51MEA
Semester	V
No. of Credits	2

Aims & Objectives of the Course

Sr. No.	Objectives
1.	To apply classical, advanced strain improvement and isolation
	techniques for fermentation processes.
2.	To optimize and sterilize media used in fermentation industry for
	commercially economical and efficient fermentations.
3.	To recover the product using suitable methods and ensuring quality of
	the finished product by quality assurance tests.
4.	To acquaint fermentation economics, process patentability, process
	validation.
5.	To understands the concepts and skills essential for industrial
	biotechnology, pharmaceuticals and bioengineering applications.

Sr. No.	Learning Outcome
1.	Students will be acquainted with the concepts of strain improvement,
	Sterilization & optimization of Media and Scale-up & Scale-down of
	fermentation
2.	Students will understand the Significance Downstream processing and
	Quality assurance of fermentation products
3.	Students will become aware about the Fermentation economics, IPR and SOPs

Syllabus Fermentation Technology– I

Unit	Title with Contents	No. of
No.		Lectures
Credit I	Upstream processes of fermentations	15
1.	 a. Concept of Fermentation Technology b. Types of fermentations: Concept of Batch, continuous and dual fermentation c. Design of a Fermenter (typical CSTR Continuous stirred tank Reactor): Different parts and their working 	2
2.	Media optimization	2
	a. Objectives of media optimization	2
	b. Methods of media optimization	
	c. Classical approach – One factor at a time, Full factorial	
	designPlackett and Burman Design (with example)	1
	d. Response Surface Methodology (RSM)	1
3.	Strain Improvement:	
	a. Objectives of strain improvement	_
	b. Methods for strain improvement	2
	 Types of mutants used in strain improvement (altered cell permeability mutants, auxotrophs, analogue resistant mutants, revertants, genetic engineering approaches) 	1
	d. Selection of different types of mutants (replica plate method, filtration enrichment, penicillin enrichment method, gradient plate technique)	
4.	Sterilization of Media:	
	a. Methods of sterilization	1
	b. Batch sterilization and Continuous sterilization (direct and	1
	indirectmethods)	1
	c. Concept and derivation of Del factor	1
	d. Filter sterilization of liquid media	1
5.	Scale-up and Scale-down:	
	a. Objectives of scale-up	1
	b. Levels of fermentation (laboratory, pilot-plant and production level-flow sheet to explain scale up)	1
	c. Criteria of scale-up for critical parameters: Scale-up windowScale-down	1
Credit II	Downstream processing and Quality assurance of fermentation	15
	products	

1.	Downstream processing of fermentation products: (method,	
	principle, types, examples of fermentations, factors affecting,	
	merits and demerits at large scale operation)	1
	a. Solid-Liquid Separation	1
	b. Cell Disruption	1
	c. Concentration d. Purification	1
	e. Formulation	1
	f. Product Recovery	1
2.	Quality assurance of fermentation products (as per IP, USP)	
	Methods of detection and Quantification of the fermentation	1
	product: physicochemical, biological and enzymatic methods	
	Bioburden test	1
	Microbial limit test	
	Sterility testing (direct inoculation method, membrane filtration	1
	method)	
	Pyrogen testing: Endotoxin detection (LAL test)	1
	Ames test and modified Ames test	1
	Toxicity testing (Acute toxicity)	1
3.	Fermentation economics, IPR and SOPs	
	Contribution of various expense heads to a process (Recurring and	1
	nonrecurring expenditures) citing any suitable example.	
	Introduction to Intellectual Property Rights – Types of IPR	1
	(Patenting in fermentation industry)	
	Concept of validation (significance of SOPs)	1

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Abeda Inamdar Senior College

Of Arts, Science and Commerce, Camp, Pune-1 (Autonomous) Affiliated to Savitribai Phule Pune University NAAC accredited 'A' Grade

T.Y.B.Sc. Microbiology

(Autonomy NEP 2023 Pattern)

Course/ Paper Title	Marine Microbiology
Course Code	23SBMB51MEB
Semester	V
No. of Credits	2

Aims & Objectives of the Course

Sr. No.	Objectives
1.	To train students in the field of Marine Microbiology
2.	To acquire advances in the knowledge of marine microbes and marine Ecology
3.	To comprehend the role of marine microbes in bioremediation
4.	To comprehend the role of marine microbes in bioprospecting.

Sr. No.	Learning Outcome
1.	The awareness of unseen and unexplored niche of marine ecosystem of
	microbes will be created in students
2.	Students will acquire advances in the knowledge of marine microbes and
	marine ecology.
3.	This course will help students to get new career opportunities in Marine
	Microbiology field.

Unit No	Title with Contents	No. of
		Lectures
Credit I		15
A	Marine ecology and sampling	
	a. Marine Habitats – estuaries, mangroves, coral reefs, salt marshes, coastal ecosystems, deep sea, hydrothermal vents, Polar habitat – Arctic, Antarctica, Southern Ocean	3
	b. Physiology of marine microorganisms – metabolic diversity, marine loop, marine snow(Composition and formation), Role of marine microorganisms in biogeochemical cycles	4
	c. Sampling methods— water sampling (Niskin sampler) and sediment sampling (Types of Grab sampler and core), Culturing methods of marine microorganisms – VBNC, biofilm, mats from vents and estuarine sample.	4
В	Applications of Marine Microbial Biotechnology	
	a. Production and applications of marine microbial products – Biosurfactants, Pigments, Enzymes, Organic acids, Antibiotics and Toxins,	2
	b. Molecular markers and their applications in fisheries and aquaculture	2
Credit II		15
A	Marine microbes, role in bioremediation and bioprospecting	
	a. Extremophilic microorganisms – econiches, different types with examples and significance	2
	b. Archaea –biodiversity, stress response, adaptation and Significance	3
	c. Marine mycology – econiche, types of marine fungi and Significance	2
	d. Bioremediation – Role of marine microorganisms in bioremediation of heavy metals, hydrocarbon pollutants – tar ball and oil spills	3
	e. Bioleaching and biodeterioration of natural and synthetic materials	2
В	Marine Microbial Diseases	
	 a. Aqua culture pathogens & Water borne pathogens - Aeromonas, Vibrio, Salmonella, Pseudomonas, Leptospira, Corynebacteria. b. Viral diseases of marine ecosystem- Effect of viruses on ecology of the marine ecosystem, movement of viruses between biomes, viral pathogens of fish, viruses in shell-fish and health hazards: Norwalk virus and Hepatitis virus A 	3

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T.Y.B.Sc. Microbiology

(Autonomy NEP 2023 Pattern)

Course/ Paper Title	Agricultural Microbiology
Course Code	23SBMB52MEA
Semester	V
No. of Credits	2

Aims & Objectives of the Course

Sr. No.	Objectives
1.	To understand plant growth improvement with respect to disease resistance, environment tolerance.
2.	To correlate stages of plant disease development, epidemiology, symptoms based classification, control methods.
3.	To understand the importance of microorganisms in sustainable agriculture, biotechnological application of bio films, edible vaccines
4.	To correlate Soil Micro biome and Role of microorganisms in soil health
5.	To determine the use of Microorganisms as tools in plant genetic engineering

Sr. No.	Learning Outcome
1.	Students will be acquainted with the concepts of plant disease resistance
2.	Students will become aware about the role of Microbiologist in Epidemiology of plant diseases
3.	Students will understand the Significance of biotechnological applications of microorganisms in the field of agriculture.

Unit No	Title with Contents	No. of
	Title with Contents	Lectures
Credit I	Plant Pathology	15
A	Stages in development of a disease Infection, invasion, colonization, dissemination of pathogens	3
В	Classification of disease based on symptoms (with one example of the following): i. Canker ii. Downy mildew iii. Mosaic	4
С	Plant disease epidemiology: i. Concepts of monocyclic, polycyclic and polyetic diseases with one example of each ii. Disease triangle iii. Forecasting of plant diseases	3
D	Methods of plant disease control i. Eradication ii. Chemical control iii. Biological control (employing bacterial and fungal cultures) iv. Integrated pest management	5
Credit II	Microorganisms in sustainable Agriculture and tools in plant genetic engineering	15
A	Microorganisms in sustainable Agriculture Soil Micro biome (plant Micro biome): Concept, Composition, functioning and methods to study plant Micro biome Conservation of soil health: Role of microorganisms in soil health	5
В	Phytonutrient availability by soil microorganisms Mechanism of diazotrophy, Phosphate solubilization, Potassium mobilization, micronutrient availability	2
С	Biofilm in plant surfaces, Biofilm formation; Biofilm in Phyllosphere and rhizosphere, Examples of plant- microbe interactions in biofilms, Biotechnological applications of plant biofilms	3
D	Microorganisms in plant genetic engineering: a. Concept of GM crops (Transgenic crops) w.r.t. to edible vaccines, insecticide resistance, herbicide resistance, improved varieties, new variants, disease resistance	2
E	Tools and techniques: i. Technology of BT resistant crops ii. Concept of edible vaccines iii. Technique of use of plant viruses in genetic engineering iv. RNAi Technology and antisense RNA technology in disease resistant plant varieties	3

References:

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M. C. E. Society's Abeda Inamdar Senior College

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T.Y.B.Sc. Microbiology (Autonomy NEP 2023 Pattern)

Course/ Paper Title	Dairy Microbiology
Course Code	23SBMB52MEB
Semester	V
No. of Credits	2

Aims & Objectives of the Course

Sr. No.	Objectives
1.	To understand prospects of dairying at commercial marketing.
2.	To acquire skills of processing of milk and dairy products
3.	To assess quality control in dairy industry.
4.	To comprehend production of dairy products of commercial significance with emphasis to local and global market demand.
	significance with emphasis to local and global market demand.

Sr. No.	Learning Outcome			
1.	Students will understand prospects of dairying at commercial marketing.			
2.	Students will acquire skills of processing of milk and dairy products.			
3.	Students will be able to assess quality control in dairy industry.			
4.	Students will be able to comprehend production of dairy products of commercial significance with emphasis to local and global market demand.			

Dairy Microbiology

Unit No	Title with Contents	No. of
		Lectures
Credit I	1. Definition, types, microflora and pathogens	10
	Definition of milk	
	Composition of milk	
	Physico-chemical properties of Milk of different animals	
	Difference between colostrum and milk.	
	Types of milk: whole, toned, double toned, homogenized, and skimmed milk, dehydrated milk	
	Microflora associated with milk and its importance.	
	Sources of contamination of raw milk and relative importance in	
	influencing quality of milk during production, collection,	
	transportation, and storage	
	Milk borne diseases.	
	2. Processing Techniques and naturally occurring preservatives	5
	Bacteriological aspects of processing techniques like	
	bactofugation, thermisation, pasteurization (in detail process is	
	expected), sterilization and boiling.	
	Naturally occurring preservative systems in milk like LP	
	system, immunoglobulins, Lysozyme, Lactoferrin etc.	
Credit II	1. Spoilage of Milk	8
	Succession of microorganisms in milk leading to spoilage	
	Stormy fermentation	
	• Ropiness	
	Sweet curdling	
	Colour and flavour defects	
	Preservation of Milk and Milk products by physical (irradiation)	
	and Chemical agents	
	• Food grade bio preservatives (GRAS)	
	Bacteriocins of LAB	

2. Microbiological asp	ects of quality control	3
a. Microbiological qua		
Dye reduction t	ests (MBRT)	
Mastitis test		
Phosphatase tes		
Microbial count	: DMC and SPC	
b. Quality assurance i	b. Quality assurance in production of milk and milk products:	
Good Manufact	uring Practices	
Sanitary standar	d operating procedures	
Total quality ma	nagement and application of HACCP program	
in dairy industr	/ .	
Safety concern of	of biofilm formation on equipment surfaces and	
their control me	asures	

References:

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Abeda Inamdar Senior College

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T. Y. B. Sc. (Under NEP 2023)

Course Title: Practical course : Immunology	Semester: V
Course Code: 23SBMB51VS	No. of Credits: 02
Nature of Course: Major	Total Teaching Hours: 60

Course Objectives		
1.	To make the students understand immunohematology techniques like crossmatching	
2.	To train the students in performing WBC differential count from peripheral blood	
4.	smear	
3.	To train the students in immunological techniques like Immunoprecipitation	
	To make the students understand the applications of recent techniques like	
4.	electrophoresis and ELISA (Antigen/ Antibody detection)	

Course Outcome		
1.	Students will learn principles of immunohematology techniques like cross matching	
2.	Students will comprehend the significance of CBC with special reference to WBC differential count from peripheral blood smear	
3.	Students will study immunological techniques like Immunoprecipitation	
4.	Students will appreciate the applications of recent immunological techniques in diagnosis of diseases	

SYLLABUS

Expt.	Topics	No. of
No.		Practicals
1.	Blood grouping: ABO and Rh system.	1
2.	Agglutination tests: Widal test (Slide test and Tube Test)	1
3.	Rapid Plasma Reagin (RPR) test	1
4.	Agglutination Inhibition (Pregnancy test)	1
5.	White blood cell differential count from peripheral blood	2
6.	Immunohematology: Cross-matching (Major and Minor)	2
7.	Coomb's test (Direct and Indirect)	1
8.	Immunochromatography tests:	2
	a) Qualitative differential detection of IgM and IgG antibodies to Dengue	
	virus in human serum / plasma	
	b) Qualitative detection of Rheumatoid factor (RA factor)	
9.	Immunoprecipitation: Double diffusion (Ouchterlony) technique	1
10.	Demonstrations of:	2
	a) Serum protein separation by electrophoresis	
	b) ELISA (Antigen/ Antibody detection)	
11.	Visit to Immunological and immune-haematological Laboratory and	1
	preparation of visit report	
	TOTAL	15

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SR. NO.	COURSE CODE	COURSE NAME
1	23SBMB61MM	Immunology - II
2	23SBMB62MM	Metabolism
3	23SBMB63MM	Genetics and Molecular Biology
4	23SBMB64MM	Practical course: Metabolism and Molecular Biology
5	23SBMB65MM	Practical course: Industrial and Food Microbiology
6	23SBMB61MEA	Fermentation Technology - II
7	23SBMB61MEB	Nanobiotechnology
8	23SBMB62MEA	Food Microbiology
9	23SBMB62MEB	Waste Management



Of Arts, Science and Commerce, Camp, Pune- 1 (Autonomous) Affiliated to Savitribai Phule Pune University NAAC accredited 'A' Grade

T.Y.B.Sc. Microbiology (Autonomy NEP 2023 Pattern)

Course/ Paper Title	Immunology II
Course Code	23SBMB61MM
Semester	VI
No. of Credits	2

Aims & Objectives of the Course

Sr. No.	Objectives
1.	To get a comprehensive understanding of Adaptive / Acquired Immunity
2.	To understand the importance of Cytokines
3.	To understand the components and mechanisms involved in Humoral Immune Response, Cell Mediated Immune Response
4.	To gain knowledge about Hypersensitivity
5.	To understands the concepts of Autoimmunity, Autoimmune diseases

Sr. No.	Learning Outcome
1.	Students will be acquainted with the concepts of Adaptive / Acquired Immunity
2.	Students will become aware about Hypersensitivity
3.	Students will understand Autoimmunity and Autoimmune diseases

Syllabus SEM VI

Unit No.	Title with Contents	No. of Lectures
Credit I	Cytokines, Adaptive / Acquired Immunity	15
1.	Cytokines:	
	Concept-Cytokines, lymphokines, monokines, interleukins,	1
	chemokines, interferons and tumor necrosis factor	_
	Properties, Attributes and biological functions of cytokines	1
2.	2. Adaptive / Acquired Immunity (Third line of defense):	
	A. Humoral Immune Response	
	i. Primary and secondary response kinetics, significance in	2
	vaccination programs	
	ii. Role of cytokines in activation and differentiation of B-cells	1
	B. Cell Mediated Immune Response	
	Activation and differentiation of T cells, role of cytokines in	2
	activation	
	Antigen processing and presentation (Major Histocompatibility class	5
	I and class II restriction pathways), cell-cell interactions and	
	adhesion molecules, response to super-antigens	
	Mechanism of Cytotoxic T lymphocytes (CTL) mediated	3
	cytotoxicity, Antibody- dependent cellular cytotoxicity (ADCC)	
	Significance of Cell Mediated Immune Response (CMI)	
Credit II	Hypersensitivity, Autoimmunity, Autoimmune diseases	15
1.	Hypersensitivity	
	General principles of different types of hypersensitivity reactions	4
	Gell and Coomb's classification of hypersensitivity – mechanism	
	with examples for type I (Immediate), II, III and IV (delayed)	
2.	Autoimmunity and Autoimmune diseases:	
	Immunological tolerance – Central and peripheral tolerance	1
	Types of autoimmune diseases	1

	Factors contributing development of autoimmune diseases	1
	Immunopathological mechanisms, Diagnosis and treatment of	4
	autoimmune diseases: Myasthenia gravis and Rheumatoid arthritis	
	Therapeutic immunosuppression for autoimmunity	1
3	Active and Passive Immunization 1.Active Immunization -Whole	2
	organism vaccines i. Attenuated vaccines ii. Inactivated vaccines iii.	
	Recombinant vaccines iv. Conjugate vaccines v. Subunit vaccine vi.	
	Toxoids	
	2.Passive Immunization Transfer of preformed antibodies 3.Latest	
	Immunization schedule in India	
4	Hybridoma Technology and Monoclonal Antibodies:	1
	Preparation, HAT selection and propagation of hybridomas secreting	
	monoclonal antibodies	
	Applications of monoclonal antibodies	

References:

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T.Y.B.Sc. Microbiology

(Autonomy NEP 2023 Pattern)

Course/ Paper Title	Metabolism
Course Code	23SBMB62MM
Semester	VI
No. of Credits	2

Aims & Objectives of the Course

Sr. No.	Objectives
1.	To learn mechanisms of transport of solutes across the membrane.
2.	To study the concept of Bioenergetics
3.	To get acquainted with mechanism of biosynthesis and degradation of Biomolecules.
4.	To comprehend basic concept of Bacterial photosynthesis

Sr. No.	Learning Outcome
1.	Students will be acquainted with the different mechanisms of transport of solutes across the membrane.
2.	Students will study the concept of Bioenergetics
3.	Students will learn mechanism of biosynthesis and degradation of biomolecules.
4.	Students will understand basic concept of Bacterial photosynthesis.

Unit No	Title with Contents	No. of lectures
Credit I	Membrane transport and Bioenergetics	15
1	Membrane transport mechanisms: i. Passive transport - Diffusion, Osmosis, Facilitated transport ii. Active transport - Active transport systems in bacteria iii. Group translocation of sugars in bacteria iv. Ionophores: Mechanism and examples	05
2	Bioenergetics: i. Laws of thermodynamics- first and second law ii. Concepts of free energy, entropy, high energy compounds: Pyrophosphate, enolic phosphates, acyl phosphates, thioester compounds, and guanidinium compounds iii. Mitochondrial electron transport chain: components, arrangement of different components in the inner membrane, structure and function of ATP	01 04 05
Credit II	synthatase, inhibitors and uncouplers of ETC and oxidative phosphorylation, energetics of mitochondrial electron transport chain Metabolic pathways and Bacterial Photosynthesis	15
1	Biosynthesis and Degradation: a. Chemistry, concept of polymerization of macromolecules: Polysaccharides. (Starch, and peptidoglycan) and Lipids (Fatty acids, triglycerides and phospholipids)	05
	b. Degradation of macromolecules – Polysaccharides (starch), Lipids (fatty acids oxidation e.g. β oxidation), Proteins (urea cycle)	05
2	Bacterial Photosynthesis: i. Examples of photosynthetic bacteria ii. Photosynthetic apparatus iii. Oxygenic and Anoxygenic mechanisms iv. Calvin cycle and its regulation	05

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T.Y.B. Sc Microbiology (Autonomy NEP 2023 Pattern)

Course/ Paper Title	Genetics and Molecular Biology
Course Code	23SBMB63MM
Semester	VI
No. of Credits	2

Aims & Objectives of the Course

Sr. No.	Objectives
1.	To get introduced to concept of recombination and bacteriophage
	Genetics
2.	To demonstrate the knowledge of common and advanced laboratory
	practices in Molecular Biology
3.	To exhibit a knowledge base in Genetics and Molecular Biology.

Sr. No.	Learning Outcome
1.	To exhibit a knowledge base in Genetics and Molecular Biology
2.	To understand the concept of cloning in bacteria
3.	To construct genetic map of bacteria and fungi by mapping techniques
4.	To get introduced to concept of recombination and bacteriophage Genetics
5.	To demonstrate the knowledge of common and advanced laboratory practices in Molecular Biology

Unit No	Title with Contents	No. of
		Lectures
Credit I	Fungal and Bacteriophage Genetics	15
	1. Fungal Genetics.	05
	a. Mendel's laws	
	b. Eukaryotic Cell cycle, Mitosis, Meiosis	
	c. Life cycle and gene mapping by Tetrad analysis in N. crassa	
	d. Life cycle and gene mapping by Parasexual cycle in A. nidulans	
	2. Bacteriophage Genetics.	10
	a. Lytic and lysogenic cycles in Virulent and	
	temperate phages. (Egs-T-series/Lamda phage)	
	b. Concept of a plaque, Phage enumeration by plaque assay.	
	c. Bacteriophage mutants: Plaque morphology (r t ype), Host	
	range and Conditional lethal mutants (Ts and Am)	
	d. Concept of Genetic Complementation, Cis-Trans test	
	e. Latest applications of Bacteriophages.	
	f. Concept of CRISPER-CAS system.	
Credit II	DNA repair mechanisms and RDT	15
	3.DNA damage and Repair mechanisms	03
	a. Concept and types of DNA damages (hydrolysis, deamination,	
	alkylation, oxidation and Radiation)	
	b. DNA repair by Photo reactivation	
	c. DNA repair by Mismatch repair mechanism	
	d. DNA repair by Excision repair mechanisms (BER/NER)	
	4. Recombinant DNA Technology-Tool sand basics of recombinant	09
	DNA technology	
	a. Introduction to recombinant DNA technology	
	b. Restriction enzymes: Nomenclature, properties and types with	
	specific examples (Eco R1, SmaI, PstI).	

c. Vectors: Features of an ideal vector	
i.Plasmids: pBR322	
ii. Bacteriophage vectors: Lambda	
iii. Cosmids	
d. Joining of DNA molecules by DNA Ligases (E. coli and T4	
phage), linkers, adaptors and homopolymer tailing.	
e. Methods to transfer recombinant DNA into bacterial host cells	
(Physical – Electroporation, Gene gun, Chemical –CaCl ₂ mediated,	
liposome mediated)	
f. Methods of screening recombinants using selectable markers and Blue-	
White screening	
5.Molecular techniques used in RDT- Principle, applications and	03
methodology of-	
a. Southern blot	
b. Northern blot	
c. Western blot	
d.PCR	

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T.Y.B.Sc. Microbiology

(Autonomy NEP 2023 Pattern)

Course/ Paper Title	Practical course: Metabolism and Molecular Biology
Course Code	23SBMB64MM
Semester	VI
No. of Credits	2

Aims & Objectives of the Course

Sr. No.	Objectives
1.	To make students aware about the clinical aspects of
	biochemistry.
2.	To make students aware about concept and preparation of buffers,
	qualitative and quantitative biochemical techniques.
3.	To make them understand basic concept of genomic DNA isolation
	and its detection.
4.	To introduce basic techniques in phage biology.

Sr. No.	Learning Outcome	
1.	Students will learn the estimation of anlysts from clinical samples by	
	biochemical techniques.	
2.	Students will learn basic concept, calculations and preparation of	
	buffers along with qualitative and quantitative biochemical techniques.	
3.	Students will be acquainted with concept, principle and methodology	
	of genomic DNA isolation and its detection using agarose gel	
	electrophoresis.	
4.	Students will understand isolation and enumeration of bacteriophages	
	from natural sources.	

Expt.	Topics	No. of
No.		Practicals
1	Clinical Biochemistry - Estimations of	4
	i) Blood sugar	
	ii) Blood urea	
	iii) Serum cholesterol	
	iv) Serum proteins	
2	Preparation of buffers	2
3	Estimation of total carbohydrates from natural sources by Phenol Sulphuric	2
	acid method.	
4	Spot tests for Proteins	1
5	Spot tests for Carbohydrates	1
6	Isolation and Enumeration of Bacteriophages (Principle,	3
	Methodology and Calculations of phage titer in PFU's/ml)	
7	Isolation of Genomic DNA from bacteria (Demonstration)	1
8	Agarose Gel Electrophoresis for DNA detection. (Demonstration)	1
	TOTAL	15



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T.Y.B.Sc. Microbiology (Autonomy NEP 2023 Pattern)

Course/ Paper Title	Practical course: Industrial and Food Microbiology
Course Code	23SBMB65MM
Semester	VI
No. of Credits	2

Aims & Objectives of the Course

Sr.No.	Objectives
1.	To make students aware about laboratory scale production of ethanol
2.	To make them understand about alcohol and sugar tolerance for yeast
3.	To introduce concept of probiotic micro flora
4.	To make them learn importance of microorganisms in food spoilage
	and use of heat in controlling microbial growth

Sr. No.	Learning Outcome
1.	Students will learn about production of ethanol at laboratory scale
2.	Students will learn to isolate probiotic micro flora and food spoilage causing organisms
3.	Students will be acquainted with significance of high temperature in controlling food spoilage
4.	Students will understand HACCP guidelines and importance of sanitary status of eatery

Expt.	Topics	No. of
No.		Practicals
1	Laboratory Scale production of the fermentation products:	3
	a. Ethanol (fermentation, recovery by simple distillation, estimation of end	
	product by CAN method and fermentation efficiency)	
2	Study of alcohol and sugar tolerance for yeast	1
3	Production and detection of amylase by shake flask or solid substrate	2
	cultivation / Bread making	
4	Isolation and identification of probiotic micro flora from natural sources or any	2
	commercial formulation.	
5	Isolation of microorganisms causing spoilage of vegetables/fruits/bread/sweets	1
6	Determination of TDP value	1
7	Determination of TDT value	1
8	Determination of D value	1
9	HACCP guidelines for food industry (activity based)	1
10	Evaluation and validation of sanitary status of an eatery	2
	i. Examination of micro flora from table surface	
	ii. Utensils	
	TOTAL	15

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T.Y.B.Sc. Microbiology

(Autonomy NEP 2023 Pattern)

Course/ Paper Title	Fermentation Technology II
Course Code	23SBMB61MEA
Semester	VI
No. of Credits	2

Aims & Objectives of the Course

Sr. No.	Objectives
1.	To understand Solid state and Submerged state fermentations
2.	To study the parameters of large scale fermentations.
3.	To acquaint with large scale fermentations of Primary Metabolites and Secondary metabolites
4.	To learn about biomass-based products in large scale fermentations
5.	To understands the concepts and skills essential for industrial biotechnology, pharmaceuticals and bioengineering applications.

Sr. No.	Learning Outcome
1.	Students will be acquainted with the concepts of Solid state and Submerged state fermentations
2.	Students will understand the Significance of large scale fermentations of Primary Metabolites and Secondary metabolites
3.	Students will become aware about the biomass-based products in large scale fermentations

Fermentation Technology-II Syllabus

Unit	Title with Contents	No. of
No.		Lectures
Credit I	Solid state and Submerged state fermentations and large scale fermentations	15
1.	Introduction to Solid State Fermentation and Submerged	
	Fermentation:	
	Process, production strains, media, fermenter design, fermentation	1
	conditions, applications, merits and demerits	
2.	Large scale production of (process with flow sheet, nature of the	
	product, production pathway, applications, production strains,	
	media, fermentation process, parameters, product recovery)	
	a. Primary Metabolites:	
	i. Vitamins (B12 and B2)	3
	ii. Amino acids - Glutamic acid, Lysine	3
	iii. Organic acids (Citric acid, Vinegar)	2
	b. Secondary metabolites:	
	i. Bioethanol	1
	Alcoholic Beverages -	2
	Beer (Lagering, Maturation, Types of beer)	
	Wine (Aging, Malo-lactic acid fermentation, types of wine, wine	
	defects, comparison of white and red wine)	
	Antibiotics [Penicillin (natural and semi synthetic) and	3
	Streptomycin]	
Credit II	Large scale production of enzymes, steroids, biomass-based	15
	products, milk products, vaccines, immune sera and Modern	
	trends in microbial production	
1.	Enzymes	
	Amylase	2
	Esterases	2
	Proteases	2

2.	Microbial transformation of steroids	2
3.	Biomass based products: i. Yeast: Baker's and Distiller's yeast ii Probiotics: Lactobacillus sporogenes	2 2
4.	Milk products: Cheese (Processed, soft, semi-hard, hard ripened types- bacterial and mold)	1
5.	Vaccines: Tetanus – Tetanus toxoid (TT) Rabies – HDCC, Chick embryo cell line, Vero cell line as per Serum Institute	1 1
6.	Immune sera: Anti-tetanus serum (ATS) Anti-rabies serum (ARS)	1 1

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Large scale production of tetanus vaccine:

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T.Y.B.Sc. Microbiology (Autonomy NEP 2023 Pattern)

Course/ Paper Title	Nanobiotechnology
Course Code	23SBMB61MEB
Semester	VI
No. of Credits	2

Aims & Objectives of the Course

Sr. No.	Objectives
1.	To study fundamentals of nanobiotechnology
2.	To train students in biogenic synthesis and characterization of Nanoparticles.
3.	To obtain understanding of various biomedical applications of nanoparticles and nanomaterials.
4.	To understand design, development and application of Nanomaterials

Sr. No.	Learning Outcome
1.	Students will get complete understanding of basic concepts in
	Nanobiotechnology
2.	Students will get theoretical knowledge about design, development and
	application of Nanoparticles and Nanomaterials.
3.	Students will get practical training inbiogenic synthesis and
	characterization of Nanoparticles. These skills will help them to apply
	for job opportunities in Nanobiotechnology field

Unit No	Topic	No. of
		Lectures
Credit I	1. Introduction to Nano-biotechnology:	15
	a. Introduction to nanoscale, nanomaterials, nanoscience and	6
	nanotechnology	
	b. Nanoscalebioassemblies -Liposomes, viruses, DNA,	
	polysaccharides and proteins (Proteinnanotubes, nanofibers,	
	peptide nanoparticles).	
	c. Biomedical applications of bioassemblies - Cell targeting, drug	
	delivery, bioimaging and vaccine development.	
	Microbial mediated metallic nanoparticles synthesis:	5
	a. Gold nanoparticles (AuNPs)	
	b. Silver nanoparticles (AgNPs)	
	c. Au-Ag alloy nanoparticles	
	d. Oxide nanoparticles	
	e. Magnetic nanoparticles	
	f. Non-magnetic oxide nanoparticles	
	g. Sulfide nanoparticles etc.	
	Characterization techniques for nanomaterials:	4
	UV-visual spectroscopy, Fourier transform infrared (FTIR), X-ray	
	diffraction	
Credit II	Applications of nanoparticles:	15
1	Antibacterial agent, drug delivery, biosensor, animal industry and nanotechnology in wastewater treatment	6
2	Microbial synthesis of metallic nanoparticle synthesis (any two)	3
3	Detection and Characterization of metallic nanoparticles in	3
	colloidal solutions by:	
	a. UV-Spectrophotometer	
	b. FTIR analysis	
		1

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T.Y.B. Sc Microbiology (Autonomy NEP 2023 Pattern)

Course/ Paper Title	Food Microbiology
Course Code	23SBMB62MEA
Semester	VI
No. of Credits	2

Aims & Objectives of the Course

Sr.	Objectives
No.	
1.	To Identify and classify types of microorganisms in food processing
	and compare their Characteristics and behavior
2.	To learn food classification based on their perishability, intrinsic and
	extrinsic factors affecting the growth of microbes in foods, role of
	microorganisms in food fermentation.
3.	To acquire knowledge about food spoilage, food borne diseases,
	predisposition and preventive and control measures

Sr. No.	Learning Outcome
1.	Students will be acquainted with the different types of microorganisms
	in food processing with respect to their classification and role.
2.	Students will learn classification of foods and different
	parameters affecting food spoilage.

3. Students will understand the importance of food borne diseases and their control measures.

Syllabus

Sr.No.	Торіс	No. of
		Lectures
Credit I	Introduction to properties of food and spoilage of	15
	food	
1	Classification of food- Perishable, non-perishable, and stable	1
	i. Definition of food	1
	ii.Sensory or organoleptic factors- appearance factors-	1
	size, shape, color, gloss, consistency, wholeness	
	iii. Textural factors-texture changes	1
	iv.Flavor factors (taste, smell, mouthfeel, temperature)	1
2	Factors affecting Microbial growth in food	
	i. Intrinsic factors- pH, water activity, O-R potential, nutrient	6
	content, biological structure of food, inhibitory substances in	
	food.	
	ii. Extrinsic factors-Temperature of storage, Relative humidity,	
	concentration of gases	
	i. Sources of food spoilage microorganisms	1
	ii. Contamination and spoilage of perishable foods-	3
	a. Vegetables and fruits	
	b. Meat and meat products	
	c. spoilage of canned foods	
Credit II	Food Preservation and food in relation to disease	15
-1	Principles of food preservation	10
1	a. Importance of TDP, TDT, D, F, Z values	
	b. Use of low and high temperature for food preservation.	
	c. Use of chemicals and antibiotics in food preservation,	
	d. Canning	
	e. Dehydration	
	f. Use of radiation	
	g. Tetra pack technology	
	h. Food grade bio preservatives	

2	Microbial food poisoning and food infection	4
	a. Food poisoning -Clostridium botulinum, Aspergillusflavus	
	b. Food infection-Salmonella typhimurium, Vibrio parahaemolyticus	
3	Concept of Prebiotic and Probiotic and fermented food-	1

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 ISBN 978-0-387- 23413-7
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Of Arts, Science and Commerce, Camp, Pune-1 (Autonomous) Affiliated to Savitribai Phule Pune University NAAC accredited 'A' Grade

T.Y.B.Sc. Microbiology (Autonomy NEP 2023 Pattern)

Course/ Paper Title	Waste Management
Course Code	23SBMB62MEB
Semester	VI
No. of Credits	2

Aims & Objectives of the Course

Sr. No.	Objectives
1.	To understand waste management and it practicable applicability.
2.	To assess the magnitude and influence of hazardous content of waste, pollution of water and waste water treatment technologies
3.	To impart the understanding of kinetics of biological systems used in waste treatment
4.	To learn the standards of waste management and competent authorities involved at National and international level.

Sr. No.	Learning Outcome	
1.	The awareness of waste management and it practicable applicability will be created in students	
2.	Students will acquire knowledge of magnitude and influence of hazardous content of waste, pollution of water and waste water treatment technologies	
3.	This course will help students to understand kinetics of biological systems used in waste treatment	
4.	Students will learn the standards of waste management and competent authorities involved at National and international level.	

Credit I A. Liquid Waste Management 15 1. Principles of Wastewater Treatment i. The need for treatment of wastewater ii. General characteristics of liquid waste - pH, Colour Turbidity, Odor, Electrical conductivity, COD, BOD, Total Solids, Total Dissolved Solids, Total Suspended Solids, Total Volatile Solids, Chlorides, Sulphates, Oil and Grease. 2. Microbiology of Wastewater Role of microorganisms in wastewater treatment i. Aerobic and Anaerobic digestion models; attached / anchored and suspended growth. ii. Removal of pathogenic microbes, indicator microbes, enumeration of different types of microbes 3. Unit operations in wastewater treatment plant i. Collection system - Methods of collection, conservancy systems, water carriage system, sewerage system. ii. Screen chamber, Grit chamber, Oil and grease removal iii. Stabilization pond, Aerated lagoon iv. Activated sludge process, Trickling filter v. Rotating biological contactors, anaerobic digestion processes, fluidized bed reactor.	Unit	Title with Contents	No. of
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fluidized bed reactor.		v. Rotating biological contactors, anaerobic digestion processes,	
		fluidized bed reactor.	

Credit II	B. Solid Waste Management and hazardous waste	15
	1. Characterization of solid wastes: Dairy and e-waste	
	2. Biomedical waste: Definition, Types, Processing	2
	3. Solid biodegradable waste processing: Composting,	2
	Vermicomposting, Biogas production	2
	4. Post-processing by-products of municipal solid waste	
	treatment: leachate refused-derived fuel (RDF)	3
2	Determination of Solids in wastewater: Total Solids, Suspended	2
	Solids, Dissolved Solids, Volatile Solids, Fixed Solids, Settleable	
	Solids	
3	Determination of Dissolved Oxygen, BOD and COD of waste	2
	water (before and after treatment) (MPCB Standards)	
4	Preparation of Project report based on a case study (Hotel/	2
	Industry-Dairy, Food processing).	
	Study of the source, generation rates and characteristics of	
	hazardous wastes and their regulation, handling, treatment,	
	and disposal. Special emphasis is placed on process design	
	of waste handling, treatment and disposal systems.	

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